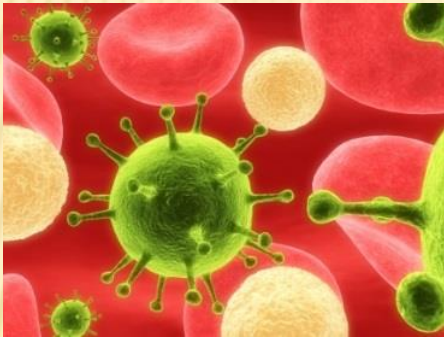




**2014 Senri Life Science International Symposium on  
“Innate Immunity, Cytokines,  
and Immune Regulation”**



**Date : January 31<sup>st</sup> (Friday), 2014 10:00-17:00**

**Venue : Senri Life Science Center Building**

**5<sup>th</sup> floor “Life Hall”**

**Coordinated by**

**Shizuo Akira & Shimon Sakaguchi**

**Immunology Frontier Research Center, Osaka University**

**SPONSORED BY SENRI LIFE SCIENCE FOUNDATION**



# 2014 Senri Life Science International Symposium “Innate Immunity, Cytokines, and Immune Regulation”

## ----- Program -----

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10:00- 10:10	Opening address	
	<b>Tadamitsu Kishimoto (President of Senri Life Science Foundation)</b>	p.2
	<b>Chair : Shimon Sakaguchi (Osaka University)</b>	
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10:10-10:50	“The non-canonical Inflammasome Pathway”	
	<b>〈Talk 1〉 Vishva M. Dixit (Genentech Inc., USA)</b>	p.4
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10:50-11:30	“Regnase-1, a ribonuclease essential for the regulation of immune responses”	
	<b>〈Talk 2〉 Shizuo Akira (Osaka University, Japan)</b>	p.8
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11:30-12:10	“Dendritic cell diversification in immune regulation”	
	<b>〈Talk 3〉 Kenneth Murphy (Washington University School of Medicine, USA)</b>	p.12
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12:10-13:10	<b>Lunch</b>	
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13:10-13:50	“Compartmentalized and systemic control of tissue immunity by commensals”	
	<b>〈Talk 4〉 Yasmine Belkaid (NIAID, NIH, USA)</b>	p.16
	<b>Chair : Shizuo Akira (Osaka University)</b>	
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13:50-14:30	“Memory T cells”	
	<b>〈Talk 5〉 Marc K. Jenkins (University of Minnesota Medical School, USA)</b>	p.20
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14:30-15:10	“Control of immune responses by regulatory T cells”	
	<b>〈Talk 6〉 Shimon Sakaguchi (Osaka University, Japan)</b>	p.24
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15:10-15:30	<b>Coffee break</b>	
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15:30-16:10	“The immune response in tuberculosis: from mouse models to human disease”	
	<b>〈Talk 7〉 Anne O’ Garra (MRC National Institute for Medical Research, UK)</b>	p.28
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16:10-16:50	“A new era for the therapy of autoimmune inflammatory diseases”	
	<b>〈Talk 8〉 Tadamitsu Kishimoto (Osaka University, Japan)</b>	p.32
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16:50-17:00	Closing remarks	
	<b>Shizuo Akira (Osaka University, Japan)</b>	p.36

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*Described time includes questions and answers.*

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## Opening Address

**Tadamitsu Kishimoto, MD.,PhD.**

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<MEMO>

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## 〈Talk 1〉

# “The non-canonical Inflammasome Pathway”

Vishva M. Dixit, MD.

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### Title:

Vice President, Research  
Genentech, Inc.  
1 DNA Way, MS40  
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### Educational History:

1982 – 1986 Postdoctoral Fellow: Department of Biological Chemistry,  
Washington University School of Medicine  
1981 – 1986 Resident: Department of Laboratory Medicine, Barnes Hospital,  
Washington University School of Medicine  
1980 – 1981 Intern: Department of Medicine, Kenyatta National Hospital  
1980 University of Nairobi, M.D.  
1975 – 1980 University of Nairobi, Nairobi, Kenya

### Research and professional experience

#### Current Appointments:

2005 – Present Vice President, Early Discovery Research, Genentech  
2006 – Present Director, Genentech Postdoctoral Program  
2006 – Present Board of Directors, Genentech Foundation  
2003 – Present Genentech Research Review Committee Member  
1999 – Present Professor, Department of Pharmaceutical Chemistry, University of California,  
San Francisco  
2011 Scientific Editor for *Cancer Discovery*

#### Past Genentech Appointments:

2003 – 2005 Vice President, Molecular Oncology  
2000 – 2003 Senior Director, Molecular Oncology  
1997 – 2000 Director, Molecular Oncology

**Past Academic Appointments:****University of Michigan Medical School**

- 1995 – 1997      Professor, Department of Pathology  
1991 – 1995      Associate Professor, Department of Pathology  
1986 – 1991      Assistant Professor, Department of Pathology

**Washington University School of Medicine**

- 1982 – 1986      Research Associate, Department of Biological Chemistry

**Awards and Honors**

- 2013    Member, National Academy of Sciences  
2012    Member, Institute of Medicine, National Academy of Sciences  
2012    Foreign Member, European Molecular Biology Organization  
2012    Member, Association of American Physicians  
2011    Member, American Academy of Arts and Sciences  
2011    Zubrod Memorial Distinguished Guest Lecturer, University of Miami, Miami, FL  
2011    HICC Distinguished Lecture, Columbia University, New York, NY  
2010    The 2010 Dan H. Campbell Memorial Lecturer  
2009    Karl Landsteiner Lecture, Vienna, Austria  
2008    Doherty Lecture, St. Jude Hospital, Memphis, TN  
2007    Daljit S. and Elaine Sarkaria Lecture, UCLA, Los Angeles, CA  
2003    Kenneth Sell Memorial lecture, Emory University, Atlanta, GA  
2000    UCSF Dean's Research Seminar, San Francisco, CA  
1999    Menten Lectureship, University of Pittsburgh, Pittsburgh, PA  
1998    Blaffer Seminar, MD Anderson Cancer Center, Houston, TX  
1998    General Motors Cancer Research Foundation, Mott Prize Selection Committee  
1997    Directors Lecture, National Institute of Health, Bethesda, MD  
1996    Hans Bloemendel Lecture, Netherlands  
1996    Member, American Society for Clinical Investigation  
1996    Warner-Lambert/Parke Davis Award in Experimental Pathology  
1983    Josiah Macy Postdoctoral Fellowship Award  
1980    Kamala Memorial Award for Best Overall Medical Student

## Abstract

Caspase-1 activation by inflammasome scaffolds comprised of intracellular nucleotide-binding oligomerization domain (NOD)-like receptors (NLR) and the adaptor ASC is believed essential for production of the pro-inflammatory cytokines interleukin (IL)- $\beta$  and IL-18 during the innate immune response. We found that C57BL/6 gene-targeted mice that caspase-11 (also known as caspase-4) is critical for caspase-1 activation and IL-1 $\beta$  production in macrophages infected with *Escherichia coli*, *Citrobacter rodentium*, or *Vibrio cholerae*. Mouse strain 129 exhibited similar defects in IL-1 $\beta$  production and harboured a mutation in the *Casp11* locus that attenuated caspase-11 expression. This finding is important because published targeting of the *Casp1* gene was done using 129 ES cells. *Casp1* and *Casp11* are too close in the genome to be segregated by recombination so the published *Casp1*<sup>-/-</sup> mice lack both caspase-11 and caspase-1. Interestingly, *Casp11*<sup>-/-</sup> macrophages secreted IL-1 $\beta$  normally in response to ATP and monosodium urate, suggesting that caspase-11 is engaged by a non-canonical inflammasome. *Casp1*<sup>-/-</sup>*Casp11*<sup>129mt/129mt</sup> macrophages expressing caspase-11 from a C57BL/6 BAC transgene failed to secrete IL-1 $\beta$  regardless of stimulus, confirming an essential role for caspase-11 in IL-1 $\beta$  production. Caspase-11 rather than caspase-1, however, was required for non-canonical inflammasome-triggered macrophage cell death, indicating that caspase-11 orchestrates both caspase-1-dependent and -independent outputs. Finally, loss of caspase-11 rather than caspase-1 protected mice from a lethal dose of lipopolysaccharide (LPS). These data highlight a unique proinflammatory role for caspase-11 in the innate immune response to clinically significant disease states.

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2. Tewari, M., Quan, L.T., O'Rourke, K., Desnoyers, S., Zheng, Z., Beidler, D.R., Poirier, G.G., Salvesen, G., and V.M. Dixit: Yama/Cpp32 $\beta$ , a Mammalian Homolog of CED-3, is a CrmA-Inhibitable Protease that Cleaves the Death Substrate Poly (ADP-Ribose) Polymerase. *Cell* Vol. 81:801-809 (1995).
3. Muzio, M., Chinnaiyan, A.M., Kischkel, F.C., O'Rourke, K., Shevchenko, A., Scaffidi, C., Bretz, J.D., Zhang, M., Ni, J., Gentz, R., Mann, M., Krammer, P.H., Peter, M.E., and V.M. Dixit: FLICE, a Novel FADD-homologous ICE/CED-3-like Protease, is Recruited to the CD95 (Fas/APO-1)



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4. Muzio, M., Ni, J., Feng, P., and V. M. Dixit: IRAK (Pelle) Family Member IRAK-2 and MyD88 as Proximal Mediators of IL-1 Signaling. **Science** Vol. 278:1612-1615 (1997).
  5. Humke, E.W., Shriver, S.K., Starovasnik, M.A., Fairbrother, W.J., and V.M. Dixit: ICEBERG: A Novel Inhibitor of Interleukin-1 $\beta$  Generation. **Cell** Vol. 103:99-111 (2000).
  6. Mariathasan, S., Newton, K., Monack, D.M., Vucic, D., French, D.M., Lee, W.P., Roose-Girma, M., Erickson, S., and V.M. Dixit: Differential activation of the inflammasome by caspase-1 adapters ASC and Ipaf. **Nature** Vol. 430:213-218 (2004).
  7. Mariathasan, S., Weiss, D.S., Newton, K., McBride, J., O'Rourke, K., Roose-Girma, M., Lee, W.P., Weinrauch, Y., Monack, D.M., and V.M. Dixit: Cryopyrin activates the inflammasome in response to toxins and ATP. **Nature** Vol. 440:228-232 (2006).
  8. Kayagaki, N., Warming, S., Lamkanfi, M., Walle, L.V., Louie, S., Dong, J., Newton, K., Qu, Y., Liu, J., Heldens, S., Zhang, J., Lee, W.P., Roose-Girma, M., and Dixit V.M. Non-canonical inflammasome activation targets caspase-11. **Nature** Vol. 479: 117–121 (2011).
  9. Qu, Y., Misaghi S., Izrael-Tomasevic, A., Newton, K., Gilmour, L. L., Lamkanfi, M., Louie, S., Kayagaki, N., Liu, J., Komuves, L., Cupp, J. E., Arnott, D., Monack, D., and Dixit V. M. Phosphorylation of NLRC4 is critical for inflammasome activation. **Nature** Vol. 490; 539-542 (2012).
  10. Kayagaki, N., Wong, M.T., Stowe, I.B., Ramani, S.R., Gonzalez, L.C., Akashi-Takamura, S., Miyake, K., Zhang, J., Lee, W.P., Muszynski, A, Forsberg, L.S., Carlson, R.W., Dixit, V.M. Non-canonical inflammasome activation by intracellular LPS independent of TLR4. **Science** Vol. 341:1246-1249 (2013).

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## 〈Talk 2〉

# “Regnase-1, a ribonuclease essential for the regulation of immune responses”

**Shizuo Akira, MD., PhD.**

---

### **Title:**

Director & Professor, WPI Immunology Frontier Research Center, Osaka University  
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### **Educational History:**

1977 M.D. Osaka University, School of Medicine  
1977-1978 Clinical Training at Osaka University Hospital  
1980-1984 Ph.D. Osaka University, School of Medicine

### **Professional History:**

1978-1980 Physician in the Department of Internal Medicine, Sakai Municipal Hospital, Sakai, Osaka  
1984-1985 Fellowship of the Japan Society for the Promotion of Science in the Institute for Molecular and Cellular Biology, Osaka University  
1985-1987 Research Fellow in the Department of Microbiology and Immunology, California University, Berkeley  
1987-1995 Research Associate in the Institute for Molecular and Cellular Biology, Osaka University  
1995- Associate Professor in the Institute for Molecular and Cellular Biology, Osaka University  
1996-1999 Professor of Biochemistry, Hyogo College of Medicine  
1996-2002 Research Head in CREST (Core Research for Evolutional Science and Technology) and SORST (Solution Oriented Research for Science and Technology) of Japan Science and Technology Corporation (JST)  
1999-present Professor of Department of Host Defense, Research Institute for Microbial Diseases, Osaka University  
2002-2007 Project Director of AKIRA Innate Immunity, ERATO (Exploratory Research for Advanced Technology) of Japan Science and Technology Corporation (JST)  
2007-present Director, WPI Immunology Frontier Research Center (WPI-IFReC), Osaka University  
2011.10-2012.3 Senior Visiting Scientist, Quantitative Biology Center, RIKEN  
2011.10-present Professor Emeritus, Hyogo College of Medicine  
2012.1-2013.3 Senior Advisor, Research Center for Allergy and Immunology, RIKEN  
2013.4-present Senior Advisor, RIKEN Center for Integrative Medical Sciences (-2014.3)  
2013.7-present Distinguished Professor, Osaka University (-2016.6)

## Honors & Awards:

- 2001 Inoue Prize for Science (Inoue Foundation of Science)
- 2001 The Hideyo Noguchi Memorial Award for Medical Sciences
- 2002 Osaka Science Prize (Osaka Science & Technology)
- 2003 Takeda Prize for Medical Science (Takeda Science Foundation)
- 2004 The Prize of Princess Takamatsu Cancer Research Fund
- 2004 Plenary Lecture TOLL2004 (First International Toll Meeting), Taormia, Italy
- 2004 Robert Koch Prize (Robert Koch Foundation, Germany)
- 2005 Medal with Purple Ribbon (Japanese Cabinet Office)
- 2006 Asahi Prize (Asahi Shinbun)
- 2006 2004-2005 "Hottest Researcher" (Thomson Scientific)
- 2006 William B. Coley Award for Distinguished Research in Basic Immunology (Cancer Research Institute, USA)
- 2007 Uehara Prize (Uehara Memorial Foundation)
- 2007 2005-2006 "Hottest Researcher" (Thomson Scientific)
- 2007 Doctor honoris causa (Technical University of Munich, Department of Medicine)
- 2007 Dunham Lectures (Harvard Medical School)
- 2007 Imperial Prize and Japan Academy Prize (Japan Academy)
- 2007 Milstein Award (International Society for Interferon and Cytokine Research)
- 2007 Thomson Research Front Award (Thomson Scientific)
- 2008 Dyer Lecture (NIH)
- 2009 Marsh Lecture (The Feinstein Institute for Medical Research)
- 2009 Keystone Symposia: Pattern Recognition Molecules and Immune Sensors of Pathogens. Keynote Lecture
- 2009 Lacey Lecture (Washington University at St. Louis)
- 2009 National Academy of Sciences of USA, Foreign Associate
- 2009 Person of Cultural Merit (Japanese Government)
- 2009 Hans Bloemendal Medal (University of Nijmegen, Netherland)
- 2010 Fellow of the American Academy of Microbiology
- 2010 17<sup>th</sup> Dundee Cell Signaling Lecture (University of Dundee, Scotland)
- 2010 Keio International Medical Science Prize (Keio University)
- 2010 Avery-Landsteiner Prize (German Society for Immunology)
- 2010 Honorary lifetime member (International Endotoxin and Innate Immunity Society)
- 2010 EMBO Associate Member
- 2010 The 2011 Canada Gairdner International Award
- 2012 The Frederik B. Bang Award (Twelfth International Conference of the International Endotoxin and Innate Immunity Society)

## Abstract

Immune responses are accompanied by dynamic changes in gene expression. Gene expression is controlled at multiple points, including signal transduction, transcription and mRNA stability. So far, transcriptional regulation has been extensively studied. Many transcription factors including NF- $\kappa$ B and AP-1 are involved in induction of genes involved in inflammatory and immune responses. However, recent studies have revealed that control of gene expression at the mRNA level is as important as transcriptional control in the immune response. Gene expression profiles obtained from human Jurkat T cells stimulated with PMA plus ionomycin revealed that regulation of mRNA stability may account for as much as 50% of all measurements of changes in total cellular polyA mRNA. We have recently identified a novel gene named Zc3h12a which has a CCCH-type zinc finger domain. The knockout mice developed spontaneous autoimmune diseases accompanied by splenomegaly and lymphadenopathy. Subsequent studies showed that Zc3h12a is a nuclease involved in destabilization of IL-6 and IL-12 mRNA via the stem loop structure present in the 3'UTR of these genes. We renamed it Regulatory RNase-1 (Regnase-1) based on the function. I would like to discuss the role of Regnase-1 in the immune response.

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## 〈Talk 3〉

# “Dendritic cell diversification in immune regulation”

**Kenneth Murphy, MD., PhD.**

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### **Title:**

Eugene Opie First Centennial Professor of Pathology & Immunology,  
Washington University School of Medicine  
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### **Educational History:**

1974-1978 BA, Summa Cum Laude, Chemistry, Rice University, Houston, TX  
1978-1984 MD, PhD, Johns Hopkins University School of Medicine, Baltimore, MD  
1984-1988 Resident-in-Training, Department of Pathology, Washington University School of Medicine, Barnes Hospital, St. Louis, MO  
1988-1989 Chief Resident, Anatomic Pathology, Washington University School of Medicine, St. Louis, MO  
1986-1990 Postdoctoral training with Dr. Dennis Y. Loh, Department of Medicine, Washington University School of Medicine, St. Louis, MO

### **Professional History:**

1989-1994 Assistant Professor, Department of Pathology, Washington University School of Medicine, St. Louis, MO  
1994-1999 Associate Professor, Department of Pathology, Washington University School of Medicine, St. Louis, MO  
1997-2003 Associate Investigator, Howard Hughes Medical Institute, Washington University School of Medicine, St. Louis, MO  
1999-Present Professor, Department of Pathology, Washington University School of Medicine, St. Louis, MO  
2003-Present Investigator, Howard Hughes Medical Institute, Washington University School of Medicine, St. Louis, MO

**Honors and Awards:**

- 1975-1977 Board of Governors Scholar, Rice University, Houston, TX
- 1977 Phi Beta Kappa
- 1978 B.A. Summa Cum Laude, Rice University, Houston, TX
- 1978 Z.W. Salsburg Memorial Award, Department of Chemistry,  
Rice University, Houston, TX
- 1978-1984 Medical Scientist Training Program Award, Johns Hopkins University School of  
Medicine, Baltimore, MD
- 1984 D.I. Macht Memorial Prize, Johns Hopkins University School of  
Medicine, Baltimore, Maryland
- 1988-1990 Juvenile Diabetes Foundation Career Development Award
- 2009 Distinguished Investigator Award, Washington University School of Medicine
- 2012 William B. Coley Award for Distinguished Research in Basic Immunology, Cancer  
Research Institute

## Abstract

We previously focused on the transcriptional basis of T-cell subset development, since these subsets coordinate the selective activation of various modules of immunity that defend against different types of pathogens. We recognized that the cytokine mediators we found to control T cell differentiation were produced by cells of the innate immune system, we next sought to understand the physiologic basis that links infection by various pathogens to the induction of an appropriate immune module. Dendritic cells (DCs) are immune lineage sensor cells responsible for detecting infection and initiating effector responses. Discovered through their capacity to activate naïve T cells, DCs also can drive effector activity in innate lymphoid cells (ILCs) by the production of mediators, activating cytokines that can also determine the type of immunity induced. Over time, DCs have been recognized as comprising several subsets defined by differing patterns of overlapping markers that may manifest distinct functions. Our work over the past 5 years has been aimed at defining the transcriptional basis of DC diversification into subsets with unique functional properties. To demonstrate the dedicated roles of these subsets *in vivo*, we developed models in which DC subsets defined by their unique transcriptional programs could be selectively deleted *in vivo*, and then analyzed the consequences of these changes for immunity to various types of pathogens. A surprising outcome of these studies has been the finding that DC subsets can exert critical and non-redundant roles in innate immune defense, in addition to their expected role in driving adaptive immune responses. The results of these types of studies suggest that DCs might have already diverged during evolution before the emergence of RAG-dependent adaptive immunity, and before they played a role in adaptive immunity. Instead, DCs may have originally served to detect pathogens of differing character and to then coordinate the selective activation of the distinct subsets of ILCs that amplify pathogen-appropriate effector modules. This could imply that the transcriptional machinery underlying DC diversification might be found in organisms lacking RAG-dependent adaptive immunity.



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## 〈Talk 4〉

# “Compartmentalized and systemic control of tissue immunity by commensals”

## Yasmine Belkaid, PhD.

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### Title:

Mucosal Immunology Section, Laboratory of Parasitic Diseases, National Institute of Allergy and Infectious Disease, NIH, Bethesda 20892, USA,

Immunology Graduate group, University of Pennsylvania, Philadelphia, PA 19104, USA

### Educational History:

- 1989 B.S., Biochemistry, University of Science & Technology Houari Boumediene of Algiers, Algeria
- 1990 M.S., with Honors, Biochemistry, University of Science & Technology Houari Boumediene of Algiers, Algeria.
- 1991 Diplome d' Etude Approfondie (DEA), Orsay University, Pasteur Institute, Paris, France
- 1996 Ph.D., with Distinction, Immunology, Orsay University, Pasteur Institute, Paris, France

### Professional history:

- 2008-Date Senior Investigator, Chief Mucosal immunology section, Laboratory of Parasitic Diseases National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland
- 2007-Date Adjunct Assistant Professor of Pathology and Laboratory Medicine, University of Pennsylvania
- 2005–2008 Head Mucosal Immunology Unit, Laboratory of Parasitic Diseases National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland
- 2002–2005 Assistant Professor, Division of Molecular Immunology  
Cincinnati Children's Hospital Medical Center, Department of Pediatrics  
University of Cincinnati, College of Medicine, Cincinnati, Ohio
- 2001–2002 Staff Scientist, Medical Entomology Section, LPD, NIAID, NIH, Bethesda, Maryland
- 1996–2001 Fogarty Fellow, Intracellular Parasite Biology Section, LPD, NIAID, NIH, Bethesda, Maryland
- 1995 Lecturer, Practical General Immunology Course  
Pasteur Institute
- 1989 World Health Organization Immunology Research and Training Center  
Department of Pathology, University Medical Center of Geneva, Switzerland
- 1988 World Health Organization Immunology Research and Training Center  
Department of Pathology, University Medical Center of Geneva, Switzerland

**Honors & Awards:**

- 2013 NIH Director's Award 2013 for groundbreaking contribution on the influence of the microbiota in host defense.
- 2012 NIAID Merit Award
- 2011 NIAID Merit Award
- 2010 NIAID Merit Award
- 2007 NIAID Merit Award
- 2003–2007 Ellison Foundation New Scholar Award in Infectious Diseases,
- 1997–1998 Phillips Foundation Award,
- 1996–1997 Phillips Foundation Award,
- 1994–1996 Foundation Marcel Merieux Award,
- 1991–1993 World Health Organization, Tropical Disease Research Fellowship,

## **Abstract**

The body is composed of various tissue microenvironments with finely tuned local immunosurveillance systems, many of which are in close apposition with distinct commensal niches. Mammals have formed an evolutionary partnership with the microbiota that is critical for metabolism, tissue development, and host defense. Despite our growing understanding of the ramifications imposed by this host-microbe alliance on immunity in the gastrointestinal (GI) tract, the degree to which individual microenvironments are controlled by resident microbiota remains unclear. We uncovered an autonomous function for the skin microbiota in controlling the local inflammatory milieu and tuning resident T lymphocyte function. Highlighting the importance of local commensal stimulation, we find that protective immunity to a cutaneous pathogen is critically dependent on skin flora and independent of gut commensals. Further, our data demonstrate that the IL-1/MyD88 pathway is essential for skin T cell function and reveal that commensals co-opt this pathway to locally tune the function of T cells. These findings underscore the importance of the flora as a distinctive feature of tissue compartmentalization and unveil unique mechanisms of immune regulation by resident commensal niches in health and disease. We will discuss the potential implications of this dialogue for the establishment of immune homeostasis, protective responses, and tissue pathology.

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## 〈Talk 5〉 “Memory T cells”

Marc K. Jenkins, Ph.D.

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**Title:**

University of Minnesota Medical School, Center for Immunology, Distinguished McKnight Professor  
Tel: 612-626-2715 Fax: 612-625-2199  
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URL: <http://www.micab.umn.edu/faculty/Jenkins.html>

**Educational History:**

1980 B.S., Microbiology, University of Minnesota  
1985 Ph.D., Immunology, Northwestern University, Mentor: Dr. Stephen D. Miller  
1985-88 Postdoctoral Fellowship, NIH, Laboratory of Immunology, Mentor: Dr. Ronald H. Schwartz

**Professional History:**

1980-81 Research Technician, Departments of Surgery and Microbiology, University of Minnesota, Minneapolis, Minnesota  
1985-87 Guest Researcher, Laboratory of Immunology, National Institute of Allergy and Infectious Diseases, NIH, Bethesda, MD  
1987-88 Staff Fellow, Laboratory of Cellular and Molecular Immunology, National Institute of Allergy and Infectious Diseases, NIH, Bethesda, MD  
1988-93 Assistant Professor, Department of Microbiology, University of Minnesota, Minneapolis, Minnesota  
1993-98 Associate Professor, Department of Microbiology, University of Minnesota, Minneapolis, Minnesota  
1997-2013 Associate Director, University of Minnesota Center for Immunology  
1998-Date Professor, Department of Microbiology, Center for Immunology, University of Minnesota, Minneapolis, Minnesota  
2013-Date Director, University of Minnesota Center for Immunology  
2013-Date President, American Association of Immunologists

**Honors and Awards:**

- 1984 Chicago Chapter Sigma Xi Research Competition
- 1989 Pew Scholars in the Biomedical Sciences Award
- 2002 American Association of Immunologists-Huang Foundation Meritorious Career Award
- 2002 Distinguished McKnight University Professor Award
- 2003 Award for Outstanding Contributions to Postbaccalaureate, Graduate, and Professional Education, University of Minnesota
- 2003 Institute for Scientific Information Highly Cited Researcher
- 2004 Academy for Excellence in Health Research, University of Minnesota Academic Health Center.
- 2008 NIH NIAID Merit Award
- 2009 1987 *Journal of Experimental Medicine* paper featured as a “Pillars of Immunology” article by the *Journal of Immunology*
- 2011 Senior Investigator Award, University of Minnesota Medical School.

## Abstract

The capacity of vertebrate hosts to clear primary infection and develop immune memory to secondary infection depends on lymphocytes that express clonally unique receptors specific for antigens from the infecting microbe. CD4<sup>+</sup> T cells express  $\alpha\beta$  T cell antigen receptors (TCRs) that recognize short microbial peptides bound to major histocompatibility complex II (MHCII) molecules on host cells, and provide protection from certain intracellular microbes. My presentation will focus on the mechanisms whereby rare naïve CD4<sup>+</sup> T cells that express TCRs specific for a given microbial peptide-MHCII ligand become activated during infection to proliferate and differentiate into effector cells, a fraction of which become memory cells.





<MEMO>

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## 〈Talk 6〉

# “Control of immune responses by regulatory T cells”

**Shimon Sakaguchi, MD., PhD.**

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### **Title:**

Professor, Immunology Frontier Research Center, Osaka University  
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### **Educational History:**

1970-1972 Kyoto University Premedical Course  
1972-1976 Kyoto University Medical School

### **Professional History:**

1976-1978 Resident, Department of Pathology, Kyoto University Medical School and Hospital  
1978-1981 Visiting Investigator, Laboratory of Pathology, Aichi Cancer Center Research Institute  
1981-1983 Senior Research Fellow, Institute for Immunology, Kyoto University Medical School  
1981-1983 Joint Appointment at the Blood Transfusion Service, Kyoto University Hospital  
1983-1985 Postdoctoral Fellow, Department of Immunology and Infectious Diseases,  
The Johns Hopkins Medical Institutions  
1986-1987 Postdoctoral Fellow, Department of Biophysics, The Johns Hopkins Medical  
Institutions  
1987-1989 Visiting Scientist, Division of Immunology and Rheumatology, Stanford University  
Medical Center  
1989-1991 Assistant Professor, Department of Immunology, Scripps Research Institute  
1992-1994 Investigator, Science and Technology Agency of Japan  
1994-1997 Head, Department of Immunopathology, Tokyo Metropolitan Institute of Gerontology  
1998-2011 Chair and Professor, Department of Experimental Pathology, Institute for Frontier  
Medical Sciences, Kyoto University  
2007-2011 Director, Institute for Frontier Medical Sciences, Kyoto University  
2011-present Professor, Department of Experimental Immunology, Immunology Frontier Research  
Center, Osaka University  
2012-present Vice-director, Immunology Frontier Research Center, Osaka University  
2013-present Osaka University Distinguished Professor

**Honors & Awards:**

- 1986 Lucille P. Markey Scholar Award in Biomedical Science
- 2003 Mochida Science Award
- 2004 Cancer Research Institute's William B. Coley Award for Basic Immunology and Tumor Immunology
- 2005 Takeda Medical Award
- 2005 Takamine Memorial Sankyo Award
- 2007 The Commendation for Science and Technology by the Minister of Education, Culture, Sports, Science and Technology, The Government of Japan
- 2008 The Keio Medical Science Prize
- 2009 Medal with Purple Ribbon
- 2012 Asahi Prize
- 2012 Japan Academy Award
- 2012 Election to Foreign Associate of the National Academy of Sciences USA

# Abstract

Naturally arising CD25<sup>+</sup>CD4<sup>+</sup> regulatory T cells (Tregs), which specifically express the transcription factor Foxp3, are engaged in the maintenance of immunological self-tolerance and immune homeostasis by suppressing aberrant or excessive immune responses, such as autoimmune disease and allergy. Key issues for understanding immunological functions of natural CD25<sup>+</sup>Foxp3<sup>+</sup>CD4<sup>+</sup> Tregs include: how they develop in the thymus and the periphery, how they suppress immune responses, and how their functional and lineage stability is established and maintained.

Assuming that Foxp3 controls the gene(s) responsible for Treg-mediated suppression, we have shown that Treg-specific deficiency of CTLA-4 results in spontaneous development of systemic lymphoproliferation, fatal T cell-mediated autoimmune disease, and hyper-production of IgE as observed in Foxp3 mutant or deficient mice. Treg-specific CTLA-4 deficiency impairs *in vivo* and *in vitro* suppressive function of Tregs, in particular Treg-mediated down-regulation of CD80 and CD86 expression on dendritic cells. In addition, the expression of CTLA-4 by developing Treg cells contributes to their acquisition of a self-skewed TCR repertoire. Thus, CTLA-4 plays critical roles in Treg-mediated suppression and their development.

Foxp3 is essential for the development of Tregs, yet its expression is insufficient for establishing the Treg cell lineage. We have recently shown that Treg development is achieved by the combination of two independent processes, *i.e.*, the expression of Foxp3 and the establishment of Treg-specific CpG hypomethylation pattern. Both are induced by TCR stimulation. The Treg-type CpG hypomethylation begins in the thymus and continues to proceed in the periphery, and can be fully established without Foxp3. The hypomethylation is required for Foxp3<sup>+</sup> T cells to acquire Treg-type gene expression, lineage stability, and full suppressive activity. Thus, those T cells in which the two events have concurrently occurred are developmentally set into the Treg cell lineage. This model explains how Treg cell fate and plasticity is controlled, and can be exploited to generate functionally stable Tregs.

How these findings can be exploited in clinical settings will be discussed.

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## 〈Talk 7〉

### “The immune response in tuberculosis: from mouse models to human disease”

Anne O' Garra, PhD., FRS, FMedSci.

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#### **Title:**

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#### **Educational History:**

1983            Graduated from MRC, National Institute for Medical Research, London,  
                         Division of Microbiology (PhD Microbial Biochemistry)  
1983-1987      Postdoctoral Fellow, Immunology, MRC, National Institute for Medical  
                         Research, London.  
1987-1989      Postdoctoral Fellow, DNAX Research Institute, Palo Alto, CA.

#### **Research Positions:**

1989-1993      Senior Research Associate, DNAX Research Institute, Palo Alto, CA.  
1993-1996      Staff Scientist, DNAX Research Institute, Palo Alto, CA.  
1996-2000      Senior Staff Scientist, DNAX Research Institute, Palo Alto, CA.  
2000-2001      Principal Staff Scientist (Director Level on the Research Track), DNAX  
                         Research Institute, CA.  
2001-            Head, Division of Immunoregulation, MRC National Institute for Medical  
                         Research, London, UK.

#### **Honors & Awards:**

2<sup>nd</sup> of Highly Cited Authors in Immunology, 1992 - 2002 (ISI Science Indicators).  
Election as Fellow of the Academy of Medical Sciences, UK - 2005.  
Election as an AAAS Fellow, in the Section on Medical Sciences  
                         (American Association for the Advancement of Science) - 2006.  
Election as Fellow of The Royal Society, UK, 2008.  
Election to EMBO membership, 2009.

#### **Memberships and Professional Activities:**

Editorial Board:  
Editor: Journal of Experimental Medicine  
Associate Editor: Immunity  
Editorial Board: Immunology

## Abstract

Tuberculosis (TB), caused by infection with *Mycobacterium tuberculosis* (*M. tuberculosis*), is a major cause of morbidity and mortality worldwide and efforts to control TB are hampered by difficulties with diagnosis, prevention and treatment. Most people infected with *M. tuberculosis* remain asymptomatic, termed latent TB, with a 10% lifetime risk of developing active TB disease, but current tests cannot identify which individuals will develop disease. We identified a whole blood transcript signature for active TB correlating with radiological extent of disease and reverting to that of healthy controls following treatment. A subset of latent TB patients had signatures similar to those in active TB patients. We also identified a distinct transcript signature that discriminated active TB from other inflammatory and infectious diseases. The immune response to *M. tuberculosis* is complex and incompletely characterized. Modular and pathway analysis of the blood transcriptome revealed that the TB signature was dominated by a neutrophil - driven interferon (IFN) - inducible gene profile, consisting of both IFN $\gamma$  and Type I IFN $\alpha\beta$  signalling. Comparison with transcriptional signatures in purified cells and flow cytometric analysis indicated that this TB signature reflects both changes in cellular composition and altered gene expression. Our studies demonstrate a hitherto under - appreciated role of Type I IFN $\alpha\beta$  signalling in human TB pathogenesis, which has implications for vaccine and therapeutic development. Our subsequent studies now provide further knowledge regarding potential mechanisms underlying the contribution of type I IFN to TB and will be discussed in depth.

The work was supported by the MRC (Grant number U117565642), and the European Research Council (European Research Council Grant ERC - 2011 - AdG 294682 - TB - PATH).

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1. Berry, M.P.R., Graham, C.M., McNab, F.W., Xu, Z., Bloch, S.J. Oni, T., Wilkinson, K.A., Banchereau, R., Skinner, J., Wilkinson, R.J.W., Quinn, C., Blankenship, D., Dhawan, R., Cush, J.J., Mejias, A., Ramilo, O., Kon, O.M., Pascual, V., Banchereau, J., Chaussabel, D., and O'Garra, A. 2010. Transcriptional Profiling Reveals an Interferon-Inducible Neutrophil-Driven Signature in Human Tuberculosis. *Nature*. 466, 973-77.
2. Redford, P., Boonstra, A., Read, S., Pitt, J., Graham, C., Stavropoulos, E., Bancroft, G., and O'Garra, A. 2010. Enhanced protection to *Mycobacterium tuberculosis* infection in IL-10-deficient mice is accompanied by an earlier and enhanced Th1 response in the lung. *Eur J*

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3. Redford PS, Murray PJ, O'Garra A. 2011. The role of IL-10 in immune regulation during M. tuberculosis infection. *Mucosal Immunol.* Mar 30.
4. McNab, F. W., Berry, M.P.R, Graham, C.M., Bloch, S.A.A., Oni, T., Wilkinson, K.A., Wilkinson, R.J.W., Kon, O.M., Banchereau, J., Chaussabel, D., and O' Garra, A. 2011. Programmed Death Ligand 1 is over-expressed by neutrophils in the blood of patients with active Tuberculosis. *Eur. J. Immunol.* Jul;41(7):1941-7.
5. Lowe DM, Redford PS, Wilkinson RJ, O'Garra A, Martineau AR. 2012. Neutrophils in tuberculosis: friend or foe? *Trends Immunol.* 33(1):14-25.
6. Pitt JM, Stavropoulos E, Redford PS, Beebe AM, Bancroft GJ, Young DB, O'Garra A. Blockade of IL-10 Signaling during Bacillus Calmette-Guerin Vaccination Enhances and Sustains Th1, Th17, and Innate Lymphoid IFN-  $\gamma$  and IL-17 Responses and Increases Protection to Mycobacterium tuberculosis Infection. *J Immunol.* 2012 Sep 12. [Epub ahead of print].
7. Bloom, CI, Graham, CM, Berry, MPR, Wilkinson, KM, Oni, T, Rozakeas, F, Xu, Z, Rossello-Urgell, J, Chaussabel, D, Banchereau, J, Pascual, J, Lipman, J, Wilkinson, RJ, and O' Garra, A. 2012. Detectable Changes in The Blood Transcriptome Are Present after Two Weeks of Antituberculosis Therapy. *PLoS One.* 7 (10), e46191.



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**〈Talk 8〉**  
**“A new era for the therapy of  
autoimmune inflammatory diseases”**  
**Tadamitsu Kishimoto, MD., PhD.**

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**Title:**

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1-4-2, Shinsenri-Higashimachi, Toyonaka-City, Osaka 560-0082, Japan  
Tel: 06-6873-2001 Fax: 06-6873-2002

**Educational History:**

1964 Graduated from Osaka University Medical School  
1970-1973 Research Fellow, Department of Medicine, Johns Hopkins University, School of  
Medicine

**Professional History:**

1973-1974 Assistant Professor in the Department of Medicine, Johns Hopkins University,  
School of Medicine  
1974-1979 Assistant Professor, Department of Medicine III, Osaka University Medical School  
1979-1983 Professor, Department of Pathology and Medicine, Osaka University Medical School  
1983-1991 Professor, Institute for Molecular and Cellular Biology, Osaka University  
1991-1998 Professor and Chairman, Department of Medicine III, Osaka University Medical School  
1995-1997 Dean, Osaka University Medical School  
1997-2003 President, Osaka University  
2003-2011 Professor of Immunology, Graduate School of Frontier Biosciences, Osaka University  
2004-2006 Member, Council for Science and Technology Policy, Cabinet Office  
2007-Present Chairman of the Board of directors, Senri Life Science Foundation  
2011-Present Guest Professor, Immunology Frontier Research Center, Osaka University

**Honors & Awards:**

- 1982 Behring-Kitasato Prize
- 1983 Osaka Science Prize
- 1986 Erwin von Bälz Prize
- 1988 Takeda Prize
- 1988 Asahi Prize
- 1990 Prize of The Japanese Medical Association
- 1990 Person of Cultural Merit, Japan
- 1991 Foreign Associate, The US National Academy of Science
- 1991 Scientific Achievement Award from the International Association of Allergology and Clinical Immunology
- 1992 Honorary Member, the American Association of Immunologists
- 1992 Imperial Prize from the Japan Academy
- 1992 Sandoz Prize for Immunology from International Union of Immunology Society
- 1992 Honorary Citizen, Tondabayashi City
- 1995 Member, the Japan Academy
- 1996 The Avery-Landsteiner Prize from the German Immunology Society
- 1997 Foreign Associate member, the Institute of Medicine of the National Academy of Science, USA
- 1997 Honorary member, the American Society of Hematology
- 1998 The Order of Culture from Emperor
- 1999 The Donald Seldin Award from the International Society of Nephrology
- 2000 ISI Citation Laureate Award
- 2001 Honorary Member, International Association of Dental Research
- 2002 Honorary Professor, the forth Military Medical University, Xi'an, China
- 2002 Honorary Member, World Innovation Foundation
- 2003 Doctor of Science, Honoris Causa, Mahidol University
- 2003 Robert Koch Gold Medal
- 2004 Clemens von Pirquet Distinguished Professor, Medicine and Immunology, University California, Davis
- 2005 Member, Deutsche Akademie der Naturforscher Leopoldina
- 2006 Honorary Lifetime Achievement Awards, International Cytokine Society
- 2009 The Crafoord Prize from the Royal Swedish Academy of Sciences
- 2010 CIS (Clinical Immunology Society, USA) President's Award
- 2011 The Japan Prize

## Abstract

A series of studies have revealed that IL-6 has a pleiotropic activity in various tissues and cells and its deregulated expression is responsible for several chronic inflammations and hemopoietic malignancies.

Humanized antibody against 80kd IL-6R (Tocilizumab) has shown significant therapeutic effect in RA, JIA, Castleman's diseases, several other autoimmune inflammatory diseases, such as progressive sclerosis, reactive arthritis, polymyalgia rheumatica, adult still's disease and uveoretinitis. Cytokine storm induced by hyperactivation of T cells has been shown to be controlled by Tocilizumab. Recently, Th17 is shown to be responsible for the pathogenesis of autoimmune diseases and IL-6 together with TGF- $\beta$  are essential for the induction of Th17.

Therapeutic effect of Tocilizumab indicates that overproduction of IL-6 is responsible for the pathogenesis of autoimmune diseases. Then, a question to be asked is how IL-6 production regulated. In a certain stimulation such as oxidized phospholipid (OxPL), overproduction of IL-6 was observed without any increase of TNF. This suggests that post-transcriptional regulation of IL-6 is important for abnormal production of IL-6. We identified a novel molecule, Arid5a, which specifically stabilizes mRNA of IL-6, sustains its overproduction and plays an important role in the promotion of these inflammatory processes and autoimmune diseases. In mice deleted of the Arid5a gene, no EAE was observed and LPS stimulation did not induce an increase of IL-6. In contrast, as was published, in Regnase-1 knockout mice, various autoimmune diseases were spontaneously induced and all mice died. Arid5a and Regnase-1 knockout mice showed mirror image phenomena regarding with autoimmune disease.

In T cells Arid5a is induced in the Th17 inducing conditions, indicating that Arid5a stabilizes not only IL-6 mRNA but also the other mRNA required for Th17 induction. Our preliminary study showed that Arid5a stabilizes STAT3 mRNA.

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## **Closing Remarks**

**Shizuo Akira, MD.,PhD.**

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